

## Evaluation of cell-mediated immune response in chickens vaccinated with Newcastle disease virus

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### Abstract

The leukocyte-migration inhibition test for the evaluation of cell-mediated immunity was developed for the use in chickens vaccinated with newcastle disease vaccine and other infected with local virulent strain of newcastle disease virus. Results indicated that the leukocyte migration test was reproducible and relatively easy assay to be performed .Serological antibody titers were determined to study the correlation between haemagglutination inhibition titers and leukocyte-migration level using the HI macro-assay .By results of this study cellular immunity level very important for evaluation the level of immunity beside the humoral immunity.

**Keywords:** Cell mediated immunity; Newcastle disease; Vaccine.

### تقييم الاستجابة المناعية الخلوية للدجاج الملقح ضد فيروس مرض النيوكاسل

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### الخلاصة

تم تقييم الاستجابة المناعية الخلوية في الأفراخ الملقحة بلقاح مرض النيوكاسل وكذلك المخمجة بعثره محلية ضارية لفايروس مرض النيوكاسل وبالاعتماد على اختبار هجرة خلايا الدم البيضاء . أشارت النتائج ان اختبار هجرة خلايا الدم البيضاء يعد قيما وسهل الإجراء وقد اعتمدت عينات المصل في دراسة العلاقة بين معايير الأضداد المثبطة للتلازن الدموي ومستوى هجرة خلايا الدم البيضاء وباستخدام اختبار تثبيط التلازن الدموي . أشارت نتائج هذه الدراسة الى أهمية دراسة المناعة الخلوية في تقييم المناعة بجانب المناعة الخلوية.

### Introduction

The Newcastle disease(ND)is an economically one of the important viral diseases(1) of poultry and its a major concern world wide (2) the sensitized lymphocytes from an immune animal when stimulated by specific antigen (3) release factors that are capable of inhibiting the migration of macrophages (macrophage inhibition factor) and blood leukocytes (leukocyte inhibition factor) (4) the level of immunity against NDV determines the severity of the disease (5).The cellular and humeral response have been

suggested to play important roles in the hosts defense against NDV infection (6,7) cell mediated immunity (CMI) has been reported as the first immunological response, being detected as early as 2-3 days after ND vaccination (7).in this study the capillary- tube leukocyte-migration inhibition test (LMI) has been used to evaluate cell mediated immunity (CMI) against ND (8), the present study presents simple under agarose LMI technique to evaluate cell- mediated immune response in chickens .

**Materials and methods**

**Chickens:** one hundred twenty one day old broiler chicks (faobro-1), were housed and kept ad libitum feed on concentrated feed. These chickens were not given any types of antibiotic or vaccine and were divided in to 4 groups, each one of 30 chicks.

First Group: thirty chicks were vaccinated at three weeks old with 0.1 ml ND vaccine orally (VG GA strain  $10^{7.1}$ EID50/0.1ml).(9)

Second Group: thirty chicks were vaccinated in three and five weeks old with 0.1 ml ND vaccine orally(VG GA strain  $10^{7.1}$ EID50/0.1ml). (10)

Third group: thirty chicks inoculated with virulent ND virus in two weeks old oronasally ( $2 \times 10^{6.5}$ EID50/0.1ml)(11).

Fourth group: thirty chicks remain as control group without vaccination

**Haemagglutination inhibition test (HI)**

Blood sample were collected from the wing vein of each bird left for clotting ,serum were separated ,the humoral antibody response was evaluated using Beta procedure(12).The highest serum dilution causing complete inhibition of haemagglutination was considered as the end point .The geometric mean HI titer for each group was expressed as the Log2 of the serum dilution(13).

**Migration inhibition test (MIT)**

Peripheral blood leukocytes of all groups were studied by capillary migration inhibition test according to (14). The migration inhibition index was calculated using the following formula:

$$\text{Migration index(MI)} =$$

$$\frac{\text{Distance of migration with antigen}}{\text{Distance of migration without antigen}} \times 100$$

Distance of migration without antigen

A 30% inhibition of migration, i.e. MI of  $\leq 70\%$  was considered as significant result, the migration index represent the mean of four reading.(15).

**Challenge test**

The vaccinated groups (one and two) were challenged with virulent strain ND virus ( $2 \times 10^{6.5}$ EID50/0.1ml) (Virology Lab , Depatment of microbiology , College of veterinary medicine , University of Mosul ) at the age of five weeks and the protection was calculated according to the percentage of mortality % ,the non vaccinated group three was challenged according to the same procedure at the age of three weeks.(16).

**Protection**

was evaluated by the calculation of % of protected chickens after challenging.

**Post mortum observation**

The gross examination changes after PM was performed on different organs (Brain , Proventriculus , Spleen , Duedenum , Trachea , Cecal tonsils) and the pathological changes were reported using semi quantity.

**Results**

The result of cell-mediated immunity by migration inhibition test gave an inhibition of migration of leukocyte with significant index and this inhibition is increased in group two more than other groups, the third group gave an inhibition of migration more than in the vaccinated group after the stimulation with ND antigen when compared with control group. Table (1).

Table 1: Result of migration indices in different groups

| Groups | Age     |         |         |         |         |
|--------|---------|---------|---------|---------|---------|
|        | 1 Day   | 3 weeks | 4 weeks | 5 weeks | 6 weeks |
| First  | 0.9 NS  | 0.8 NS  | 0.5 S   | 0.45 S  | 0.40 S  |
| Second | 0.87 NS | 0.83 NS | 0.63 S  | 0.56 S  | 0.43 S  |
| Third  | 0.85 NS | 0.85 NS | 0.40 S  | 0.40 S  | 0.38 S  |
| Fourth | 0.88 NS | 0.87 NS | 0.85 NS | 0.80 NS | 0.80 NS |

S= Significant      NS=Non Significant

The titer of antibody which was measured by haemagglutination inhibition test showed a significant increase in titer of the vaccinated groups but this increase was more remarkable in third group while the infected group presented highest titer after inoculated with virulent strain. Table (2). The challenge with virulent strain virus gave highest mortality in the non vaccinated group , the animals of

this group present after PM severe gross lesions in different organs , while the vaccinated group gave high protection rate with mild changes after post mortum, in vaccinated group when compare with one vaccination, the protection rate was higher in boosted animal group (Table 3).

Table 2: Variation of haemagglutination inhibition test titer in different groups.

| Group NO. | Age      |             |            |            |           |
|-----------|----------|-------------|------------|------------|-----------|
|           | 1 Day    | Three weeks | Four weeks | Five weeks | Six weeks |
| First     | 2Log 2   | 1Log 2      | 3.8Log 2   | 3.1Log 2   | 2.7Log 2  |
| Second    | 2.2Log 2 | 1.1Log 2    | 4Log 2     | 3.7Log 2   | 5.5Log 2  |
| Third     | 1.9Log 2 | 1.2Log 2    | 4.7Log 2   | 6.2Log 2   | 5.6Log 2  |
| Fourth    | 2.2Log 2 | 1Log 2      | 0Log 2     | 0Log 2     | 0Log 2    |

Table 3: the protection rate and mortality with post mortum lesion In different groups

| Group No. | Organs |                |        |          |         |               | % mortality rate | % Protection rate |
|-----------|--------|----------------|--------|----------|---------|---------------|------------------|-------------------|
|           | Brain  | Proventriculus | Spleen | Duodenum | Trachea | Cecal tonsils |                  |                   |
| First     | +      | ++             | ++     | +        | +       | ++            | 10/30            | 66.9%             |
| Second    | —      | —              | +      | —        | +       | +             | 4/30             | 86.6%             |
| Third     | ++     | —              | ++     | ++       | ++      | +++           | 17/30            | 56.6%             |
| Fourth    | —      | —              | —      | —        | —       | —             | —                | —                 |

( — ) = no changes  
 ( + ) = mild congestion

( ++ ) = moderate congestion with enlargement  
 ( +++ ) = severe inflammation with bleeding

**Discussion**

Various approach have been used for identifying the specific components of the immune system involved in protection.

Results of present study indicate that the cellular immune response of chicken vaccinated with ND vaccine can be evaluated by the migration inhibition test, this result is an agreement with that of (3,15). The leukocyte migration inhibition test does not require any expensive

equipment and advantage of this technique is the plates can be fixed , stained and evaluated at any time and / or save as a permanent record . Although an inverted microscope was used to measure the migration area in the present study. A ruler could have sufficed (3,8) so cell mediated immunity (CMI) is been suggested to be an important factor in the development of protection in chicken vaccinated with ND vaccine (11,12,13).

The humoral immunity was proved to be protected too by HI antibodies against ND (3).

The objective of this study was to ascertain whether (humoral and cellular) immunity are key components in the protection of chicken against ND (12,13). Therefore vaccination programs should be directed toward eliciting and maintaining high immunity level to NDV in flock of bird also the test of the migration inhibition could be used to evaluate cell mediated immunity against other infectious disease, in addition to ND (8,16).

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#### **References**

1. Parede L, Young P L. The pathogenesis of velogenic Newcastle disease virus infection of chickens of different ages and different level of immunity. *Avian disease*.1990; 39: 803-808.
2. Aini I. Endogenous chickens production in South East Asia. *Wild Poultry Sci J*.1990; 46: 51-57.
3. Reynolds DL, Maraqa A D. Protection immunity against newcastle disease: The role of cell mediated immunity *avian Disease*.2000; 44:145-159.
4. Young D F, Randell R E , Lawrenson J A , Souberbielle B E. Clearance of a persistent paramyxovirus infection mediated by cellular immune response but not by serum neutralization antibody. *J Virol*.1990 ; 64: 5403-5411.
5. Reynolds D L, Maraqa A D. Protection immunity against Newcastle disease :The role of antibodies specific to Newcastle disease virus poly peptides. *Avian Disease* 2000; 44:138-144..
6. Beard C W , Brugh A. Immunity to Newcastle disease. *Am J Vet Res*. 1975 ; 509-512.
7. Andreason C B , Latimer K S. Separation of avian heterophils from blood using Ficoll- Hypaque discontinuous gradients *Avian disease*.1989; 33:163-167.
8. Timms L. Leukocyte migration inhibition as an indicator of cell mediated immunity in chickens. *Avian Pathol*.1974; 3:174-187.
9. Vlaovic M S , Buening G M , Coan R W. Capillary tube leukocyte Migration Inhibition as a correlate of cell mediated immunity in chickens. *Cell Immunol*.1975; 17:335-341.
10. Wehmann E , Herczeg J , Tanyi J , Nagy E , Commiczi B. Lentogenic field isolates of Newcastle disease virus isolated in Canada and Hungary are identical with vaccine type used in region. *Avian Pathology*.1999;28:6-12.
11. Nasser M , Lohr J E , Mebrata G Y , Zessin K H , Baumann M P O , Adem Z. Oral newcastle disease vaccination trial in Ethiopia. *Avian Pathol*.2000 ; 27-37.
12. Alexander D J , Manvell D J. Experimental assessment of the pathogenicity of the newcastle disease viruses outbreaks in Great Britain in 1997 for chickens and turkeys and the protection afforded by vaccination. *Avian Pathol*.1999 ; 28: 501-511.
13. Steel D B , Torrie J H. Principles and procedures of statistics. 2<sup>nd</sup> ed., McGraw-Hill Company ,Inc., London. 1980.
14. Maas R A , Oei H L , Kemper S , Koch G , Visser L. The use of homologous virus in the Haemagglutination –inhibition Assay after vaccination with Newcastle disease virus strain Lasota or Clone 30 leads to an over estimation of protective serum antibody serum. *Avian Pathol*.1998 ; 27:625-627
15. Pawan K A , Reynold D L. Evaluation of the cell mediated immune response of chickens vaccinated with newcastle disease virus as determined by the under –agarose leukocyte migration inhibition Technique. *Avian disease*. 1991; 35:360-364.
16. Biswas H R , Haque M M , Oxley M , Prodhan M A M. A comparative study on the protection of indigenous chickens against Newcastle disease include Australian NDV4 HR and locally produced conventional vaccines in Bangladesh. *Prev Vet Med*.1996 ; 26(2) : 157-164.